

A diagram of a gel filtration chromatography column. It is a vertical tube with a blue background, containing a light blue gel matrix with circular pores. Red and green dots representing molecules are shown at different levels within the pores. The column has a white inlet at the top and a white outlet at the bottom. White dotted lines extend horizontally from the left and right sides of the column.

# GEL FILTRATION CHROMATOGRAPHY

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# INTRODUCTION

Gel Filtration is a separation technique which is strictly based on molecular size. It is a helpful method to separate the macromolecule of different sizes from one another. It is also known as Gel permeation, molecular, size exclusion or molecular sieve chromatography. Water is a mobile phase in Gel filtration chromatography whereas organic solvent used as a mobile phase in case of Gel permeation Chromatography.



# GENERAL PRINCIPLES

Gel filtration chromatography is a special case of partition chromatography because the stationary phase is the same as the mobile phase. The stationary phase is in form of porous

when a sample containing solutes varying from small to large molecules elutes through column. Small molecules penetrate all pores and are retained, thus being eluted later than large molecules, which move only in mobile phase. Molecules of intermediate size penetrate only some pores, thereby being retained to a lesser degree than small ones. The elution of the solute is best characterized by a distribution coefficient,  $K_d$ .

$$K_d = \frac{V_o - V_e}{V_s}$$

$V_o$  = Volume of solvent required to elute an unretained compound; also called "void volume".

$V_e$  = Volume of the mobile phase required to elute a compound from the chromatographic column.

$V_s$  = Volume of the stationary phase.

The Distribution Coefficient.

$$K_{av} = \frac{V_e - V_o}{V_t - V_o}$$

- $V_t$  = Volume of the chromatographic column

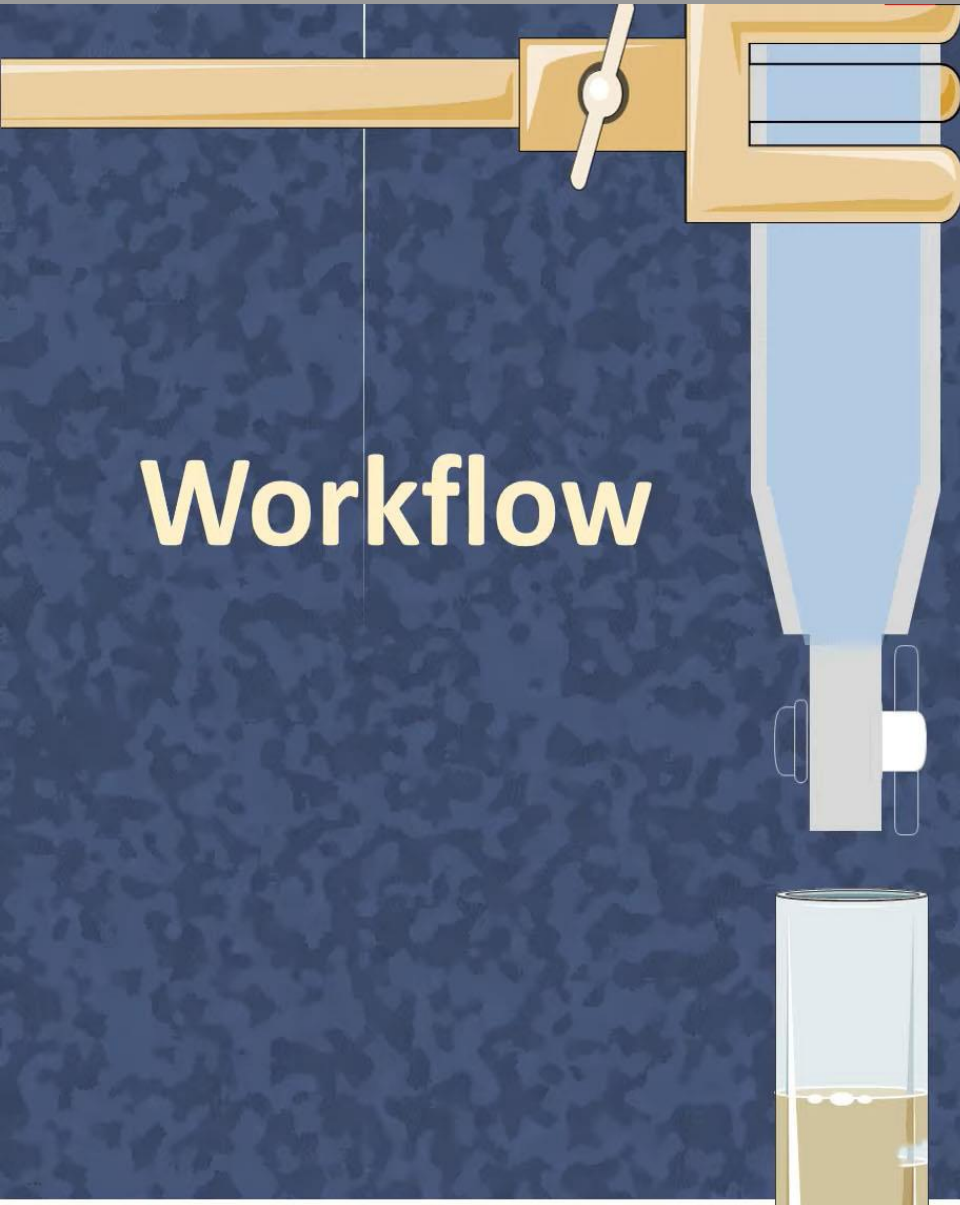
# Workflow

Equilibration

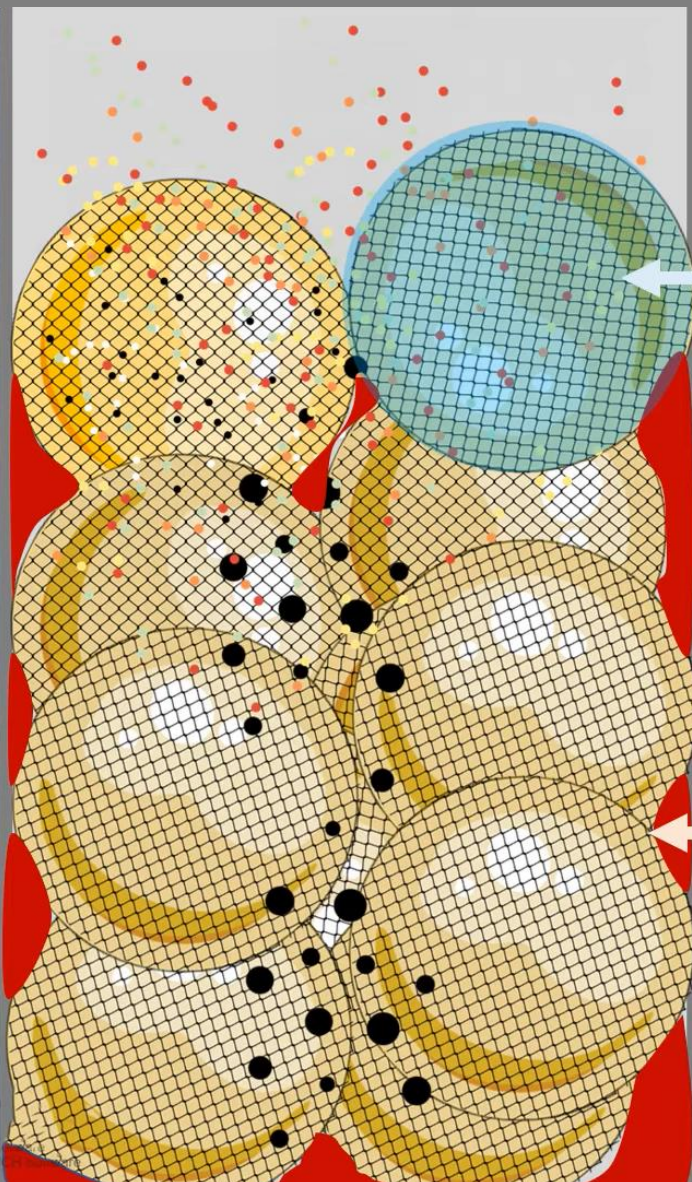
Binding

wash

Elution







**Internal volume**

$V_i$

**Void volume**

$V_o$

# TYPES OF MEDIA

The best-known media for gel-filtration are based on dextran, a polymer of glucose cross-linked with epichlorohydrin. These are called Sephadex. The degree of swelling is determined by the degree of cross-linking.

(i) Sephadex: The Sephadex types are designated by G, followed by a number which corresponds to ten times the water regains, the number of millimeters of water taken by 1 gm of dry gel matrix. Thus, the commercially available series: Sephadex G-10, G-15, G-25, G-50, G-100, G-200 have fractionation ranges for proteins of molecular weights up to 600,000 Daltons.

Sephadex also swells in organic solvents such as dimethyl sulphoxide, formamide, dimethyl formamide and mixtures of lower alcohols with water. However, the degree of swelling in these organic solvents or their mixtures is less than in water alone. The Sephadex series, LH-20, LH-60 and Sephacryl are suitable for organic solvents. SEPHADEX LH types are specially prepared for gel filtration in organic solvents. These are prepared by the hydroxy- propagation of Sephadex G-50. Which have both hydrophilic and lipophobic properties.

(ii) Sepharose: This gel is based on agarose, it is used mostly for the fractionation of large molecules. The agarose base is obtained by the purification of agar devoid of charged polysaccharides. The polysaccharide chains in agarose consist of D-galactose and 3,6 anhydro-L-galactose forming double helices like DNA which aggregate on ageing to form bundles. These bundles are mechanically strong with pore size capable of fractionating macromolecules of different molecular weight.

(iii) Sephacryl: This is an example of a mixed gel, (agarose and poly- acrylamide). The agarose forms a support frame for the polyacrylamide gel. It is prepared by covalently cross-linking allyl dextran with N' N' - methylene bis acrylamide to give a rigid gel with a carefully controlled range of pore size.



S No.	Types of Sephadex	Fractionation range for Peptides and globular proteins, MW(Daltons)	Water Regain Gm Water/gm Dry Gel	Bed Volume Cm./gm. Dry Gel	Types of Bio-gel	Fractionation Range (Dalton)
1	G-10	Up to 700	1.0	2.0	P -2	100-1800
2	G-15	Up to 1500	1.5	3.0	P -4	800-4000
3	G-25	1000-5000	2.5	5.0	P -6	1000-6000
4	G-50	1500-30000	5.0	10.0	P -10	1500-20000
5	G-75	3000-70000	7.5	12-15	P -30	2500-40000
6	G-100	4000-150000	10.0	15-20	P -60	3000-60000
7	G-150	5000-450000	15.0	12-30	P- 100	5000-100000
8	G-200	5000-800000	20.0	30-40	P- 150	15000-150000

Type	Approximate Agarose Concentration In %(w/v)	Approximate Exclusion Mol. Wt.(Daltons)	
Sepharrowse-2B	2	Polysaccharides	Protein
		20 X 10 <sup>6</sup>	40 X 10 <sup>6</sup>
Sepharrowse-4B	4	5 X 10 <sup>6</sup>	20 X 10 <sup>6</sup>
Sepharrowse-6B	6	1 X 10 <sup>6</sup>	4 X 10 <sup>6</sup>

# CHARACTERISTICS OF GEL MATRIX

The characteristics of such gels are enumerated below:

- (a) Chemically inert: The gel matrix should be inert. Any chemical interaction between the matrix and solutes may lead to irreversible binding to the gel material and chemical alterations of labile substances.
- (b) Stability: The matrix must be stable over a wide range of pH and temperature.
- (c) Controlled particle size and size distribution: The particle-size must possess good flow rate characteristics. Therefore, it is essential that the particle size distribution be as narrow as possible.
- (d) Mechanically rigid: This is essential for the gel particles, so as to prevent deformation by the forces caused by the flow of liquid (solvent)

# APPLICATIONS OF GEL FILTRATION CHROMATOGRAPHY

There are many different applications of gel permeation chromatography. These have been employed in biomedical research problems and clinical laboratory. These include:

- (i) Separation of polysaccharides, enzymes, antibodies and other proteins.
- (ii) Separation of nonpolar species such as triglycerides in non-aqueous mobile phases.
- (iii) Desalting process i.e., removal of salt and buffer ions from protein solutions as an alternative to dialysis.
- (iv) Phenol removal from nucleic acid preparations.
- (v) Removal of products, viz., co-factors and inhibitors from enzymes.

# *bibliography*

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- J.A. Dean ,Chemical Separation Methods , New York
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